Supplementary Material for manuscript

“Harmful algae and export production collapse in the equatorial Atlantic during the zenith of Middle Eocene Climatic Optimum warmth”

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1 Supplementary Methods

1.1 Palynology
We use the palynological materials from Cramwinckel et al., (2018). Tablets containing a known amount of *Lycopodium clavatum* spores were added prior to palynological processing to allow for quantification of the absolute number of dinocysts per sample. Samples were then treated with 30% HCl and ~38–40% HF and isolation of the 15–250 µm fraction was established using nylon mesh sieves and an ultrasonic bath to break up agglutinated particles of the residue. Palynomorphs were counted up to a minimum of 200 identified dinocysts. Dinocyst taxonomy as cited in (Williams et al., 2017) was followed. Functional ecological dinocyst grouping follows (Brinkhuis, 1994; Sluijs et al., 2005; Frieling and Sluijs, 2018). In particular, the Protoperidinioids, cysts of Protoperidiniaceae, are thought to derive from heterotrophic dinoflagellates and proliferate under increased nutrient conditions (Sluijs et al., 2005). Goniodomids (cf. Sluijs and Brinkhuis, 2009), cysts of Goniodomaceae, including *Polysphaeridium* spp., are considered to be characteristic for restricted, typically lagoonal settings, and can tolerate large swings in salinity (Brinkhuis, 1994; Frieling and Sluijs, 2018). Organic linings of benthic foraminifera were also incorporated into the palynological counts, the presence of which indicates presence of benthic foraminifera.

1.2 Core images
For characterization of bioturbation and color changes throughout the MECO interval, high resolution images of Cores 13R–23R (archive halves) were made using a GEOTEK Geoscan-III linescan camera (aperture 6.7 and 8.0) at MARUM, Bremen University. Changes in sedimentary structures were assessed by scoring the types of bioturbation and by measuring the maximum burrow diameter in each section. Burrows were measured from the digital core photos. Presence of trace fossils in the sediment indicates presence of metazoa living at the seafloor. Food supply is a major factor governing body size of benthic organisms (Danovaro et al., 2014; Rex et al., 2006; Wei et al., 2010). Therefore, burrow size can be used as a qualitative indicator for downward organic carbon fluxes.

1.3 Bulk carbonate stable isotope ratios
Stable carbon (δ¹³C) and oxygen (δ¹⁸O) isotope ratios of bulk carbonate (40 samples; 0.3–2 mg of freeze-dried and powdered sample) were measured at the Royal Netherlands Institute for Sea Research (NIOZ), using a Thermo Scientific Kiel IV carbonate device coupled to a Thermo Scientific MAT 253 isotope ratio mass spectrometer. Values are reported relative to the Vienna Pee Dee Belemnite (VPDB) standard. Analytical (internal) precision is ±0.07‰ for δ¹³C and ±0.1‰ for δ¹⁸O, supported by NBS-19 and internal laboratory (VICS) standards.

1.4 Organic carbon and nitrogen content, organic carbon isotopes
Samples were analyzed for total organic carbon (TOC) and nitrogen contents, as well as the carbon isotopic composition of TOC (δ¹³C_{org}). ~0.3 g of freeze-dried bulk sediment was powdered and decalcified using 1 M
HCl. Samples were dried in a stove at 50°C, and subsequently, TOC and N contents were measured on ~10 mg of powdered, homogenized residue using a CNS analyzer (Fisons). Stable carbon isotope ratios were determined using an isotope ratio mass spectrometer (IRMS; Finnigan DELTAplus) coupled online to the CNS analyzer. Absolute reproducibility, based on international and in-house standards, for TOC and δ¹³Corg was better than 0.1% and 0.05‰, respectively. TOC content of the sediment is primarily governed by organic matter flux to the sediment and bottom water oxygen conditions. Both molar C/N ratios and δ¹³Corg provide indications of organic matter sourcing (e.g., Meyers, 1994).

1.5 Bulk sediment elemental composition

Major and minor element composition of the sediment (37 samples) was measured. As the first step in acid digestion, ~125 mg of freeze-dried and powdered sample was dissolved in a mixture of 2.5 mL HClO₄:HNO₃ 3:2 and 2.5 mL HF (40%) in closed polytetrafluoroethylene autoclaves at 90 ºC overnight. Acids were evaporated at 160 ºC and the resulting residue was re-dissolved in 25 mL 4.5% HNO₃ at 90 ºC overnight. Elemental compositions of these total acid digested solutions were measured at Utrecht University using Inductively Coupled Plasma-Optical Emission Spectrometry (ICP-OES; Perkin Elmer Optima 3000). Analytical uncertainty was better than 5% based on duplicates and in-house standards. Most of the measured Mo concentrations were below 20 ppm and thus below the background emission line. The error on these measurements can be >10% and values should therefore be regarded as approximates. Under oxygen-depleted bottom-water conditions, several elements are actively scavenged from the water column into the sediments. Elements such as V, Cr, Cu, Zn, Ni and in particular Mo get enriched in the sediment under such conditions, and their sedimentary contents thus provide a proxy for bottom-water paleo-redox conditions (Brumsack, 1980; Tribovillard et al., 2006; Scott and Lyons, 2012). Detrital elements such as Al, Ti, K and Fe can provide evidence for detrital sediment providence. Element abundances were assessed in absolute concentrations, but also normalized to Aluminium (Al), in order to correct authigenic enrichment for dilution by the biogenic components of the sediment - here predominantly biogenic silica. Following recommendations by Van der Weijden (2002), regression analysis for normalized elemental values was omitted.

1.6 Organic geochemistry

As fully described in Cramwinckel et al. (2018), organic compounds were extracted from Site 959 sediments using an Accelerated Solvent Extractor (ASE) and separated into apolar, polar and neutral fractions using column chromatography. Here, we desulphurized five of the polar fractions across the MECO interval using Raney nickel. These were analysed by gas chromatography-mass spectrometry (GC-MS) at the Royal Netherlands Institute for Sea Research (NIOZ) to assess the presence of isorenieratene and its derivatives. The compound isorenieratene is a photosynthetic pigment that derives solely from green sulphur bacteria (Overmann et al., 1992). Presence of this biomarker or its derivatives thus indicates development of photic zone euxinia (Sinninghe Damsté et al., 1993), although absence of preserved isorenieratene in the sediment is not evidence of absence of photic zone euxinia.
1.7 Stratigraphy

Core recovery in the middle part of the MECO is poor (Figure 2). Peak MECO temperatures were recorded in Sections 959D-19R-1 and 959D-18R-5. Recovery of Core 959D-19R is poor (7.6%) and the position of Core 959D-19R relative to -18R and -20R is uncertain, as we were unable to precisely correlate core data from this interval to downhole logging data (Mascle et al., 1996). This raises the question whether the recorded MECO warming in available core material underestimates the total warming that occurred at Site 959. Based on the following lines of evidence, we argue that, while the 3.5°C temperature change remains a minimum estimate, it is unlikely a gross underestimate. First, based on the evolution of MECO temperatures as recorded in other sites (Bohaty et al., 2009; Bohaty and Zachos, 2003), peak temperatures are typically reached at the end of the event. As we record similar temperatures in the top of Core 959D-19R and bottom of 959D-18R, it is likely that both of these core intervals represent the interval of peak MECO warming. Our complementary δ¹⁸O record (Figure DR1) also shows minimum values at the end of the MECO (upper Core 959D-18R), supporting interpreted peak warmth. Secondly, the rapid warming within Core 959D-19R itself also suggests this core interval sampled the peak of the event. Therefore, we argue that our record most likely captures close to the full extent of the SST change, and we can thus make inferences about peak MECO relative to MECO warming and cooling. In this, we refer to the phase of warming based on TEX₈₆ SST as “MECO warming”, the phase of peak MECO SST as “peak MECO”, and the SST decrease back to pre-MECO SSTs as “MECO recovery”.


2 Supplementary Figures

Figure DR1. Additional geochemical and palynological records of oceanographic and environmental change during the MECO at ODP Site 959. All against depth in meters below seafloor (mbsf). Plotted with recovery of cores in black. A. Oxygen isotopic composition ($\delta^{18}$O) of bulk carbonate. B. TEX$_{86}$H-based sea surface temperature (SST) reconstruction (Cramwinckel et al., 2018). Yellow shading represents elevated MECO temperatures, with peak MECO temperatures in orange shading. C. Branched and Isoprenoid Tetraether (BIT) index. D. Absolute quantitative abundances of terrestrial palynomorphs (grains/gram). E. Absolute quantitative abundances of dinocysts (cysts/gram). F. Absolute quantitative abundances of organic linings of benthic foraminifera (linings/gram). G. Molar $C_{org}/N_{bulk}$ ratio. H. Carbon isotopic composition ($\delta^{13}$C) of bulk organic material (per mille; ‰). I. Manganese (Mn) contents in ppm (solid lines) and normalized to Al (dotted lines).
Figure DR2. Light microscope images of the dinocyst *Polysphaeridium zoharyi*. A, B. Specimen from 959D-18R-4W, 15-17 cm. C. Specimen from 959D-18R-5W, 85-87 cm. D. Multiple specimens from 959D-18R-5W, 85-87 cm. Scale bars represent 25 um. A–C share the same scale.
3 Supplementary Tables

Table DR1. Presence (black infill) and absence (white infill) of bioturbation types and maximum burrow diameter (mm) present in ODP Hole 959D, core 13R-23R, for all sections longer than 70 cm. The two sections in which the highest MECO SSTs and lowest average burrow diameter (Figure 2) were found are outlined in red.
Table DR2. Summary of inferred (export) productivity change during the MECO.

<table>
<thead>
<tr>
<th>Ocean basin</th>
<th>Region</th>
<th>Site / Section</th>
<th>Proxy / Proxies</th>
<th>MECO phase</th>
<th>Signal</th>
<th>Interpretation</th>
<th>References</th>
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<td>Atlantic</td>
<td>equatorial</td>
<td>ODP Site 959</td>
<td>TOC%, trace elements, dinocysts, bioturbation</td>
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<td>-</td>
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<td>2 (peak warmth)</td>
<td>Polyplaxaedurum peak, bioturbation size decrease</td>
<td>decreased export productivity</td>
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<td>3 (recovery)</td>
<td>Redox-sensitive trace element increase, TOC% increase</td>
<td>decreased bottom-water oxygenation, possibly increased export productivity</td>
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<td>northwest</td>
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<td>Siliceous microfossils, benthic foraminifera</td>
<td>1</td>
<td>Siliceous microfossil flux increase</td>
<td>increased export productivity</td>
<td>(Witkowski et al., 2014; Moebius et al., 2015)</td>
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<tr>
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<td>Siliceous microfossil flux decrease, planktic and benthic foram flux increase</td>
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<td>decreased export productivity</td>
<td>(Boscolo-Galazzo et al., 2015, 2014)</td>
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<td>2</td>
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<td>Alano</td>
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<td>Eutrophic planktic and planktonic foram assemblages, nannofossil assemblages somewhat more eutrophic</td>
<td>increased export productivity</td>
<td>(Boscolo-Galazzo et al., 2013; Luciani et al., 2010; Spofforth et al., 2010; Toftman et al., 2011)</td>
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<td>3</td>
<td>Eutrophic planktic and planktonic foram assemblages, TOC%-rich intervals</td>
<td>even more increased export productivity</td>
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<td>(Lyle, 2005)</td>
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<td>decreased productivity</td>
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4 Supplementary References


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